

Stereospecific Analysis of Fatty Acid Composition of Chufa (*Cyperus esculentus* L.) Tuber Oil

Moonjung Kim · Siwon No · Suk Hoo Yoon

Received: 2 July 2007 / Revised: 23 August 2007 / Accepted: 30 August 2007 / Published online: 15 September 2007
© AOCS 2007

Sir,

Chufa (*Cyperus esculentus* L.) is a sedge-like plant known as earth almond, nut grass, or yellow nut sedge. The chufa tuber is regarded as a novel food lipid resource due to its substantial oil content, almond-like flavor, and ease of cultivation [1, 2]. Chufa tubers contain 17–25% oil on a dry weight basis. Some of the physicochemical characteristics of chufa tuber oil are similar to those of olive oil. Chufa tuber oil could be a potential new source of stable edible oil. The fatty acid composition of the triacylglycerols and the polar lipids of chufa tuber oil have been reported elsewhere, and oleic acid was the major fatty acid reported in both lipid classes [3, 4].

The chemical, physical and biological characteristics of lipids are largely dependent upon the composition and positional distribution of fatty acids on the glycerol backbone, thus the stereospecific analysis of fatty acids in the triacylglycerol was considered important for use of the lipid for both dietary and industrial purposes [5, 6]. It has been reported that the major fatty acid present at the sn-2 position of triacylglycerols of chufa tuber oil was oleic acid, similar to olive oil, although no information was available regarding the fatty acid composition of chufa tuber oil at the sn-1 and sn-3 positions [1].

We determined the positional fatty acid distribution of chufa tuber at stereospecific positions; the sn-1, sn-2, and sn-3 positions in the triacylglycerols of chufa tuber oil.

Chufa tubers were washed with water, and oven dried to a moisture content of 20%. The total lipid (TL) was extracted in a Soxhlet apparatus using a mixture of chloroform and methanol at 40 °C. The total lipid extracted was dissolved in chloroform and separated into neutral lipids (NL), glycolipids (GL) and phospholipids (PL) using silicic acid column chromatography [7]. Stereospecific analysis of chufa oil was carried out using pancreatic lipase, phospholipase A₂, TLC and GC [8]. The fatty acid composition was determined using gas chromatography [9]. The fatty composition at the sn-3 position was calculated based on the equation, $3 \times [\text{TG}] - [\text{sn-1} + \text{sn-2 position}]$.

The total lipid content of the chufa tuber was 17.6% on a dry weight basis. The neutral lipid (90% of the total) was the main lipid component in the chufa tuber oil, while glycolipid and phospholipid were present at 6.9 and 3.1%, respectively. The major fatty acids of the chufa tuber oil were identified as oleic (65.6%), linoleic (16.2%), palmitic (15.4%), and stearic (2.2%) acids in decreasing order (Table 1), while the minor acids were linolenic and palmitoleic. The oleic acid content of chufa tuber oil is much greater than that of most other vegetable oils, such as sunflower oil (23.6%), soybean oil (24.9%), or corn oil (23.8%), but comparable to that of olive oil [10–13]. Chufa tuber oil with its high percentage of oleic acid, should be relatively stable and resistant to oxidation [1, 3].

The fatty acid composition of the NL fraction was similar to the TL; however, the concentrations were slightly different. The NL contained slightly more oleic acid and slightly less palmitic and linoleic acid. The GL contained more palmitic, linoleic, and linolenic acid and

M. Kim
Department of Food Science and Nutrition,
Inha University, Incheon, South Korea

S. No · S. H. Yoon (✉)
Food Functionality Research Center,
Korea Food Research Institute,
San 46-1 Baekhyundong, Bundanggu,
Seongnamsi, Kyunggido 463-746, South Korea
e-mail: shyoon@kfri.re.kr

Table 1 Fatty acid composition of chufa tuber lipids

| | Relative content (%) | | | | | |
|---------------|----------------------|-------------------|-------------------|-------------------|-------------------|-------------------|
| | C _{16:0} | C _{16:1} | C _{18:0} | C _{18:1} | C _{18:2} | C _{18:3} |
| Total lipid | 15.4 | 0.2 | 2.2 | 65.5 | 16.2 | 0.5 |
| Neutral lipid | 14.6 | 0.1 | 2.3 | 68.3 | 14.4 | 0.3 |
| Glycolipid | 20.4 | – | 2.1 | 58.3 | 18.5 | 0.7 |
| Phospholipid | 24.3 | – | – | 53.4 | 22.3 | – |

Table 2 Positional fatty acid distribution of chufa tuber oil

| Oil | sn-position | Relative content (%) | | | | | |
|--------------------|-------------|----------------------|-------------------|-------------------|-------------------|-------------------|-------------------|
| | | C _{16:0} | C _{16:1} | C _{18:0} | C _{18:1} | C _{18:2} | C _{18:3} |
| Chufa tuber | 1 | 12.8 | – | 3.3 | 60.4 | 23.5 | – |
| | 2 | 0.9 | 0.1 | 0.2 | 75.1 | 23.1 | 0.6 |
| | 3 | 32.6 | 0.4 | 3.1 | 61.1 | 1.9 | 0.9 |
| Olive ^a | 1 | 13.1 | 0.9 | 2.6 | 71.8 | 9.8 | 0.6 |
| | 2 | 1.4 | 0.7 | – | 82.9 | 14.0 | 0.8 |
| | 3 | 16.9 | 0.8 | 4.2 | 73.9 | 5.1 | 1.3 |

^a Reference [13]

had less oleic acid than the NL. Fatty acids associated with sugar moieties are very unsaturated in higher plant chloroplasts [14]. The PL fraction contained greater amounts of palmitic and linoleic acid and lower amounts of stearic, oleic, and linolenic acid compared to the other lipid classes in chufa tuber oil.

The positional fatty acid distribution of chufa tuber oil is shown in Table 2. Oleic acid was the major fatty acid in the sn-1 and sn-2 positions. The percentage composition of fatty acids in the sn-1 and sn-2 positions was nearly the same as that of the total fatty acid composition; however, the concentration was slightly different. Oleic acid was also the main fatty acid in the sn-3 position followed by palmitic, stearic, linoleic, linolenic, and palmitoleic acid. This differed from others who reported that the fatty acids located at the sn-3 position was highly influenced by their respective concentration in the total fatty acids at the other positions [8, 15, 16]. Palmitic acid and stearic acid were distributed primarily in the sn-1 and sn-3 positions, while oleic acid and linoleic acid were found in greater amounts at the sn-2 position. In most vegetable oils, unsaturated fatty acids occupy the sn-2 position and saturated fatty acids are located in the sn-1 and sn-3 positions. The

preponderance of linoleic acid at the sn-2 position is well known in vegetable oils [5, 8, 16]. The distribution of fatty acids between sn-1 and sn-3 position was nearly symmetrical for palmitic acid, stearic acid, and oleic acid, which is in agreement with Brockerhoff and Yurkowski [8]. Palmitoleic acid and linolenic acid were primarily associated with the sn-3 position, with undetectable amounts in the sn-1 position.

References

1. Linssen JPH, Kielman GM, Cozijnsen JL, Pilnik W (1988) Composition of chufa and olive oils. *Food Chem* 28:279–285
2. Linssen JPH, Cozijnsen JL, Pilnik W (1989) Chufa (*Cyperus esculentus*): a new source of dietary fiber. *J Sci Food Agric* 49:291–296
3. Oderinde RA, Tairu OA (1988) Evaluation of the properties of yellow nutsedge (*Cyperus esculentus*) tuber oil. *Food Chem* 28:233–237
4. Oderinde RA, Tairu OA (1992) Determination of the triglyceride, phospholipids and unsaponifiable fractions of yellow nutsedge oil. *Food Chem* 45:279–282
5. Hunter JE (2001) Studies on effects of dietary fatty acids as related to their position on triglycerides. *Lipids* 36:655–668
6. Yoon SH, Kim E (2003) Recent progress in enzymatic production of structured lipids. *Food Sci Biotechnol* 12:721–726
7. Sahasrabudhe MR, Chapman DG (1961) Partial fractionation of fatty acid triglycerides on a silicic acid column. *J Am Oil Chem Soc* 38:88–92
8. Brockerhoff H, Yurkowski M (1966) Stereospecific analyses of several vegetable fats. *J Lipid Res* 7:62–64
9. Jung MO, Ju JW, Choi DS, Yoon SH, Jung MY (2002) CLA formation in oils during hydrogenation process as affected by catalyst types, catalyst contents, hydrogen pressure, and oil species. *J Am Oil Chem Soc* 79:501–510
10. Warner K, Knowlton S (1997) Frying quality and oxidative stability of high-oleic corn oils. *J Am Oil Chem Soc* 74:1317–1322
11. Romero A, Cuesta C, Sanchez-Muniz FJ (1998) Effect of oil replenishment during deep-fat frying of frozen foods in sunflower oil and high-oleic acid sunflower oil. *J Am Oil Chem Soc* 75:161–167
12. Chung J, Choe E (2001) Effects of sesame oil in thermooxidative stability of soybean oil. *Food Sci Biotechnol* 10:446–450
13. Kiritsakis AK (1990) Chemistry of olive oil. In: Kiritsakis AD (ed) Olive oil. AOCS, Champaign, pp 25–55
14. Gunstone FD, Norris FA (1983) Lipids. In: Lipids in foods. Rergamon, Oxford, pp 8–14
15. Roche IA, Weber EJ, Alexander DE (1971) Effects of fatty acid concentration and positional specificity on maize triglyceride structure. *Lipids* 6:531–536
16. Sempore G, Bezar J (1991) Determination of molecular species of oil triglycerols by reversed-phase and chiral-phase high-performance liquid chromatography. *J Am Oil Chem Soc* 68:702–709